## BUKTI KOREPONDENSI

JURNAL ILMIAH
JURNAL INTERNASIONAL BEREPUTASI DAN BERFAKTOR DAMPAK WoS IF > 1,0 (SYARAT KHUSUS)

| NO | JENIS ISIAN | ISIAN |
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| 1 | Judul Artikel | Cis-2 and trans-2-eicosenoic fatty acids are novel inhibitors for Mycobacterium tuberculosis protein tyrosine phosphatase A |
| 2 | Penulis | 1. Lalu Rudyat Telly Savalas; 2. Baiq Repika Nurul Furqon; 3. Dina Asnawati; 4. Jannatin 'Ardhuha; 5. Prapti Sedijani; 6. Saprizal Hadisaputra; 7. Baiq Nila Sari Ningsih; 8. Jufrizal Syahri |
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| 14 | Apakah ini syarat khusus | Ya (First author dan corresponding author, faktor dampak > 1,0) |
| 15 | Keterangan (Opsional) | Memenuhi syarat khusus faktor dampak > 1,0 <br> (https://mjl.clarivate.com/journal-profile) (akun MJL institusional dibutuhkan untuk melihat informasi detil jurnal dimaksud menurut pengindeks Clarivate Analytics), atau <br> https://ojs.ptbioch.edu.pl/index.php/abp/about, atau sumber lain: https://academic-accelerator.com/Impact-Factor-IF/Acta-Biochimica-Polonica |

## KRONOLOGI KOREPONDENSI

| No | Tanggal | Aktivitas | Keterangan |
| :---: | :---: | :---: | :---: |
| 1 | 10 Maret 2020 | Submission acknowledgement | Email dari Editor Acta Biochimica Polonica (ABP) |
| 2 | 10 Maret 2020 | Permintaan melengkapi submission | Diskusi di OJS ABP |
| 3 | 11 Maret 2020 | Tambahan kelengkapan submsission | Via OJS ABP |
| 4 | 11 Maret 2020 | Tambahan kelengkapan submission | Email ke ABP |
| 5 | 20 April 2020 | Diskusi proses review | Saran reviewer potensial |
| 6 | 5 Juni 2020 | Acceptance dengan revisi minor | Diskusi di OJS, konfirmasi acceptance telah dikirim via email tetapi tidak masuk mailbox author. Acceptance dan revisi disampaikan via OJS. |
| 7 | 5 Juni 2020 | Tanggapan rerhadap permintaan revisi minor | Diskusi di OJS |
| 8 | 5 Juni 2020 | Cover letter revisi manuskrip | Dikirim author ke OJS ABP |
| 9 | 6 Juni 2020 | Full manuskrip setelah revisi | Dikirim author ke OJS ABP |
| 10 | 8 Juni 2020 | Copyedit | Dikirim author ke OJS ABP |
| 11 | 14 Juni 2020 | Permintaan proofreading author | Diskusi via email |
| 12 | 15 Juni 2020 | Jawaban author terhadap permintaan proofreading akhir | Via email |

Dokumen di bawah ini disusun menurut kronologi seperti pada tabel di atas.

## K1. Submission Acknowledgment



K2. Permintaan melengkapi metadata

## [ABP] New Submission without Metadata completed 5201

## Participants

Mrs. Malgorzata Goraj-Basaj (mgb)
Dr hab. Katarzyna Potrykus (kpotrykus)
Lalu Rudyat Telly Savalas (savalas123)

## Messages

| Note | From |
| :--- | :--- |
| Lalu Rudyat Telly Savalas: | mgb |
| Dear Authors, | $2020-03-10$ |

Thank you for the submission 5201 entitled: "Cis-2 and trans-2-eisocenoic fatty acids are novel inhibitors for Mycobacterium tuberculosis Protein tyrosine phosphatase A," in Acta Biochimica Polonica.

However, according to Instructions for Authors three names with affiliations and mailing addresses of potential reviewers should be added as comments.

You are also asked to send the e-mails of ALL the Authors as according to the rules of COPE all Authors must be notified about the progress of the submission.

Without this step manuscript could not be processed.
Sincerely
Małgorzata Basaj

Mrs. Malgorzata Goraj-Basaj
Polish Biochemical Society, Warsaw
abp@ptbioch.edu.pl

## K3. Tambahan kelengkapan submission

- Thank you for your response.

I noted the instruction but missed the steps during submission in the new OJS system. Can I just add the requested potential reviewers as words file attached to this presubmission menu? Also, for the the email address of the co-authors, how can I edit and add them via the system? (I would suggest that the email addresses of co-authors denoted as compulsary during submission process, so that the same problem can be avoided in the future. Alhthouh I understand that it is stated in the guidelines).

Thank you and I will be back to fullfil the request.
Kind regards,

LR Telly Savalas

The e-mails please can be added at this step only by me so write them in the message.
As to the reviewers you can attached them as word file also in the message not tin the
mgb
2020-03-11 09:02
AM submission menu.

Sincerely

Małgorzata Basaj

- Thank you. It is noted.

I just send you the requested information via email.
savalas123
2020-03-11 02:09
PM

I look forward for a positive outcome.

Kind regards,
Telly
Recommended reviewers for submission 5201

- Dessy Natalia, Institut Teknologi Bandung, Indonesia, dessy@chem.itb.ac.id
mgb
2020-03-11 10:11 PM
- Carlos Polanco, Universidad Nacional Autónoma de México, E-mail: polanco@unam.mx
- Magdalena Druszczyńska, University of Lodz, E-mail: majur@biol.uni.lodz.p

K4.


K5. Diskusi proses review

## [ABP] A message regarding Acta Biochimica Polonica 5201

## Participants

Mrs. Malgorzata Goraj-Basaj (mgb)
Dr hab. Katarzyna Potrykus (kpotrykus)
Lalu Rudyat Telly Savalas (savalas123)

## Messages

## Note

From

Dear Dr Savalas, mgb
We have trouble in finding a willing person for commenting your paper.

Could you kindly send us three names with mailing addresses and affiliations?

It would speed up the reviewing process.

Sincerely
Małogrzata Basaj

The following message is being delivered on behalf of Acta Biochimica Polonica.

- Dear Ms Basaj, thank you for your email. Herewith I recommend potential reviewers for my manuscript: 1. Dr. Roil Bilad, Biochemical Process, Universiti Petronas Malaysia. Email: mroil.bilad@utp.edu.my 2. Prof. Iman Permana Maksum, Dept of Chemisty, University Padjajaran Bandung Indonesia. Email: ip_maksum@unpad.ac.id 3. Dr. Jaspreet Kaur Dhanjal, National Institute of Advance Science, Japan, Email:
jaspreetk.dhanjal@aist.go.jp 4. Dr. Ni Nyoman Sri Budayanti, Udayana University, Bali, Email: nyomansribudayanti@gmail.com I hope it helps the process. Kind regards, Telly


## K6. Acceptance dengan perbaikan minor

Dear Telly,
I've send the message with the decision to your email address through the system,
but maybe it ended up in your spam folder (?). Anyway, I am attaching the message AM
below,
Cheers,
Kasia

From: "Dr hab. Katarzyna Potrykus" [k.potrykus@abp.ptbioch.edu.pl](mailto:k.potrykus@abp.ptbioch.edu.pl)

To: "Lalu Rudyat Telly Savalas" [telly@uram.ac.id](mailto:telly@uram.ac.id), "Baiq Repika Nurul Furqon" [refika.nf007@gmail.com](mailto:refika.nf007@gmail.com), "Dina Asnawati" [dinaasnawati@gmail.com](mailto:dinaasnawati@gmail.com), "Jannatin 'Ardhuha" [j.ardhuha@unram.ac.id](mailto:j.ardhuha@unram.ac.id), "Prapti Sedijani" [praptisedijani@unram.ac.id](mailto:praptisedijani@unram.ac.id), "Saprizal Hadisaputra" [rizal@unram.ac.id](mailto:rizal@unram.ac.id), "Baiq Nila Sari Ningsih" [6110220044@email.psu.ac.th](mailto:6110220044@email.psu.ac.th), "Jufrizal Syahri" [jsyachri@umri.ac.id](mailto:jsyachri@umri.ac.id)

## Subject: [ABP] Editor Decision reference number 5201

reference number 5201 entitled: "Cis-2 and trans-2-eisocenoic fatty acids are novel inhibitors for Mycobacterium tuberculosis Protein tyrosine phosphatase A"

Dr Lalu Rudyat Telly Savalas, Baiq Repika Nurul Furqon, Dina Asnawati, Jannatin 'Ardhuha, Prapti Sedijani, Saprizal Hadisaputra, Baiq Nila Sari Ningsih, Jufrizal Syahri Dear Dr Lalu Rudyat Telly Savalas, Baiq Repika Nurul Furqon, Dina Asnawati, Jannatin 'Ardhuha, Prapti Sedijani, Saprizal Hadisaputra, Baiq Nila Sari Ningsih, Jufrizal Syahri

## I am happy to inform you that your paper can be accepted for publication in Acta

 Biochimica Polonica after minor revisions.Please find the comments below this letter.

Please consider carefully the report and amend the manuscript accordingly. You are also kindly requested to answer in detail to all comments and describe them in the accompanying letter which should be attached as the first page of the revised manuscript.
When preparing the revised version of your manuscript please refer to the Instructions to Authors and to the latest issue of Acta Biochimica Polonica available on-line at https://ojs.ptbioch.edu.pl/index.php/abp/about/submissions to follow the style and requirements of the journal.

It is assumed that the final manuscript will be accepted by all the authors.

Please send the revised version of the article (with changes marked in color) and the cover letter describing the changes made to the manuscript by the system (under Revisions).

Thank you in advance for your prompt revision, Sincerely

Dr hab. Katarzyna Potrykus
University of Gdańsk, Gdańsk, Poland
k.potrykus@abp.ptbioch.edu.pl

## Reviewer D:

The new idea of verification of potential inhibitors of protein tyrosine phosphatase (PtpA) of Mycobacterium tuberculosis is sound and reasonable. The authors extended this attracting and straightforward approach to test the potential inhibitors such as cis-2 and trans-2 eicosenoic fatty acids. The calculated IC In values as 11.26, 8.20 and 27.97 mM for trans-2, cis-2 and trans-11-eicosenoic fatty acid, clearly indicates it potential. Additionally, docking analysis of PtpA inhibition by cis-2-eicosenoic acid is identyfying four aminoacids $\mathrm{Asn}_{14}, \mathrm{Ile}_{15}, \mathrm{Cys}_{16}, \mathrm{Arg}_{17}$ as binding with strong interaction with cDocker energy -37,1939 kcal/mol. This constitutes the goals and novelty of the manuscript.

On the other minor note the following suggestion/correction should be incorporated into the text before publication:

1. 2. Page 5 , line 15 the word "was" should be replaced with "is"
1. 2. Page 6 . line 18 the words "In agreement with inhibition assay" should be moved and incorporated at the end of the sentence.
1. 2. Page 8. line 14. The sentence should be corrected by insertion "that" before "rather" and by inserting at the end of the sentence the words "strongly interact with PtpA"

The manuscript is of good quality and importance and is written in order to meet the standard for the articles published in Acta Polonica Biochimica. I certainly recommend it for publication in the current version with incorporation of these small changes.

Recommendation: Revisions Required

The following message is being delivered on behalf of Acta Biochimica Polonica.

- Dear Dr. Potrykus, savalas123
thank you for you notification. I have attached the requested revision as well as a cover letter for the revision.

I hope that the raised issues are solved.
Should there is furhter concern, please do not hesitate to contact me.
Kind regards,
Telly
P.S. It seems that I got trouble in receiving the requested revision. I cannot find it in spam box as well, but thank you that this OJS platform works well.

K7. Respons terhadap permintaan revisi minor

## Copyediting file submission 5201

## Participants Edit

Mrs. Malgorzata Goraj-Basaj (mgb)
Dr hab. Katarzyna Potrykus (kpotrykus)
Lalu Rudyat Telly Savalas (savalas123)

## Messages

Note
From

Please find our Words file for submission 5201.
As requested, we have omitted markup to allow further

Thank you.

Kind regards,
Dr. Lalu Rudyat Telly Savalassavalas123, Novel Mtb PtpA Inhibitor submitted to Act
Biochim Polonica LRT Savalas submission 5201
Copyediting.docx

## K8. Cover letter revisi manuskrip

## Dear ABP Editor,

Thank you for the requested revision for submission 5201. In respoding to the requested revision, herewith I list the revision we made:

1. Issue 1: Page 5, line 15 the word "was" should be replaced with "is"

Original text:
"Lysate of untransformed E. coli BL21was used as control in each experiments."

Replaced with:
"Lysate of untransformed E. coli BL21 is used as control in each experiments."
2. Issue 2: Page 6. line 18 the words "In agreement with inhibition assay" should be moved and incorporated at the end of the sentence.

## Original text:

"In agreement with inhibition assay, docking of PtpA with trans-11-eicosenoic acid prone to have weaker interaction in term of both energy and the distance of amino acid residues."

## Replaced with:

"Docking of PtpA with trans-11-eicosenoic acid prone to have weaker interaction in terms of both energy and the distance of amino acid residues which is in agreement with the inhibition assay."
3. Issue 3: Page 8. line 14. The sentence should be corrected by insertion "that" before "rather" and by inserting at the end of the sentence the words "strongly interact with PtpA"

## Original text:

"It is somehow surprising that a small molecule such as eicosenoic acid can strongly interact with PtpA, as many researchers have reported rather complex inhibitors."

Replaced with:
"It is somehow surprising that a small molecule such as eicosenoic acid that rather complex inhibitors as many researchers have reported strongly interacts with PtpA."

To conclude this letter, I have the opinion that the requested revision has been properly made. I also confirm that the revised manuscript refers to the latest Author Guidelines.
Thank you and I look forward to hearing from you.

Kind regards,
Dr. Lalu Rudyat Telly Savalas
Dept of Chemistty, Faculty of Teacher Training and Education
University of Mataram
Jl. Majapahit No. 62 Mataram
Nusa Tenggara Barat, Indonesia 83125
Email telly@unram.ac.id; rudyat_telly@yahoo.com

## K9. Manuskrip yang telah direvisi

# Cis-2 and trans-2-eicosenoic fatty acids are novel inhibitors for Mycobacterium tuberculosis protein tyrosine phosphatase A 

Lalu Rudyat Telly Savalas ${ }^{1 *}$, Baiq Repika Nurul Furqan², Dina Asnawati ${ }^{2}$, Jannatin<br>${ }^{\prime}$ Ardhuha ${ }^{3}$, Prapti Sedijani ${ }^{4}$ Saprizal Hadisaputra ${ }^{1}$, Baiq Nila Sari Ningsih ${ }^{1,5}$, Jufrizal Syahri ${ }^{6}$<br>${ }^{1}$ Department of Chemistry Education, Faculty of Teacher Training and Education, University of Mataram, Indonesia; telly @unram.ac.id<br>${ }^{2}$ Department of Chemistry, Faculty of Mathematics and Natural Sciences, University of Mataram;<br>${ }^{3}$ Department of Physics Education, Faculty of Teacher Training and Education, University of Mataram;<br>${ }^{4}$ Department of Biology Education, Faculty of Teacher Training and Education, University of Mataram;<br>5 Department of Chemistry, Faculty of Science, Prince of Songkhla University, Thailand<br>${ }^{6}$ Department of Chemistry, Universitas Muhammadiyah Riau, Indonesia

* Correspondence: telly@unram.ac.id; Tel.: (+62 (0370) 623873)


#### Abstract

Small protein tyrosine phosphatase (PtpA) of Mycobacterium tuberculosis (Mtb) is attributed to the development of latent tuberculosis infections, and hence becomes an interesting target for drug development. In this communication, inhibition of PtpA by naturally occurring fatty acids: cis-2 and trans-2-eicosenoic acids is investigated. Mtb PtpA was heterologously expressed in Escherichia coli, and the activity of PtpA was inhibited by cis-2 and trans-2 eicosenoic fatty acids. Both compounds showed strong inhibition of $\operatorname{PtpA}$ activity, with $\mathrm{IC}_{50}$ at low micromolar concentrations. In comparison, trans-11-eicosenoic acid only slightly inhibited PtpA. In silico analysis confirmed the inhibition of PtpBA by cis-2-eicosenoic acid by formation of several hydrogen bonds. These findings show that cis-2 and trans-2 eicosenoic fatty acids are potential candidates for latent tuberculosis inhibitors.


Keywords: Mycobacterium tuberculosis, latent infection, Pprotein tyrosine phosphatase A, cis-2 and trans-2 eicosenoic fatty acids

## 1. Introduction

Mycobacterium tuberculosis pathogen is one of the most devastating pathogens with high mortality all over the world. The death toll caused by this infection is worsened by increasing HIV infection cases. This figure is even more complicated by the development of antibiotic resistant strains: multi-drug resistant tuberculosis (MDR TB) and extensively drugresistant TB (XDR TB). The battle against TB has an additional front, i.e. latent TB infection (LTBI). It is estimated that 1.7 billions of people, or approximately one quarter of the world's population, are infected with these bacteria in the latent infection fashion (World Health Organization, 2019; Houben \& Dodd, 2016), and $10 \%$ of those individuals develop active infection at a later stage of their life (Vynnycky \& Fine, 2000; Stutz et al., 2018).

The ability of the bacteria to avoid the acidic lysosomal degradation within host macrophages has been recognized as one of the survival mechanisms of $M$. tuberculosis from phagolysosome degradation, and leads the bacterium to its latent phase (Pieters \& Gatfield, 2002), where it further utilizes nutrition from its host for long term dormancy (Mali \& Meena, 2018). Although several bacterial survival factors, such as the protein tyrosine phosphatases, bacterial lipoarabinomannan (LAM) and protein kinase $G$ (PknG) have been extensively suggested to be involved in the latent infection (Li \& Xie, 2011; Janssen et al., 2012), the mechanism by which latent TB infection develops into its active state is not fully understood. Nevertheless, inhibition of latency factors opens up the research field to combat bacteria even before latency is established.

Among the proteins associated with the development of latent TB infection is protein tyrosine phosphatase A (PtpA). This protein is known to be responsible for inhibition of fusion even between phagosome and lysosome. In a normal endocytosis pathway, once the bacterium is engulfed by the macrophage cell, the resulted mycobacterial-laden phagosome recruits host vacuolar- $\mathrm{H}^{+}$-ATPase (V-ATPase) that acidifies the endosome, a precondition
required later when it fuses with the lysosome and thus ensuring that a suitable environment for hydrolytic enzymes of the lysosome is formed (Sun-Wada et al., 2009; Stutz et al., 2018; Upadhyay et al., 2018). However, bacterium develops a survival strategy by secreting PtpA that permeates the phagosome membrane and binds to the subunit H of V -ATPase in the macrophage cytosol. PtpA is also reported to dephosphorylate vacuolar sorting protein VPS33B which is required for the fusion of endocytic organelles. The binding of subunit H of V-ATPase and dephosphorylation of VPS33B by mycobacterial PptA are concerted evens that account for the hinderance of bacteria to enter phagolysosome degradation (Bach et al., 2008; Wong et al., 2011; Korb et al., 2016). PtpA was also reported to supress the host innate immunity by regulating host gene expression (Wang et al., 2017). From this point of view, PtpA, along with other effector proteins, becomes an interesting target for anti-latent TB drug development.

Several attempts to discover novel drugs benefitted from the progress in the computer aided drug discovery (CAD). A recent example of this approach has been underlined by Zhang and co-workers who reported a thiobarbiturate compound as a novel Mtb PtpB inhibitor with an $\mathrm{IC}_{50}$ of $22.4 \mu \mathrm{M}$ (Zhang et al., 2019). Furthermore, a thiosemicarbazone compound predicted by molecular modelling has been synthesized and revealed its inhibitory effect on Mtb PtpA with a low micromolar $\mathrm{IC}_{50}$ (Sens et al., 2018). A comprehesive review of in silico studies targetting tubercular protein is presented by de Oliveira Viana et al. (de Oliveira Viana et al., 2018). A recent in silico study by Dhanjal and co-workers has suggested that eicosenoic fatty acid derivative, the trans-2 eicosenoic acid. might inhibit Mtb PtpA and a related phosphatase of Mtb, PtpB (Dhanjal et al., 2014). The present study aims at testing the ability of trans-2-eicosenoic fatty acid to inhibit Mtb PtpA.

## 2. Materials and Methods

### 2.1 Materials

## Plasmid and Escherichia coli strains

Recombinant plasmid pET30b-PtpA was a kind gift from Prof. Yossef Av-Gay, Univesity of British Columbia, Canada. All-The plasmids wasere maintained in Escherichia coli XL1-Blue, and for expression, E. coli BL21(DE3) was used. Bacteria were grown in LB medium containing $0.5 \%$ yeast extract, $1 \% \mathrm{NaCl}, 1 \%$ bacto trypton and $30 \mu \mathrm{~g} / \mathrm{mL}$ kanamycin (USP Biobasic). Agar LB medium was made by addition of $2 \%$ bacto agar. All ingredients were from major biochemical vendors.

## Chemicals

All reagents for buffers were from major chemical vendors. Cis-2-eicosenoic and trans-2-eicosenoic fatty acids were purchased from Larodan AB (Sweden). Trans-11-eicosenoic fatty acid and para-nitrophenyl phosphate were from Sigma-Aldrich.

### 2.2. Methods

### 2.2.1 Expression of Mtb protein tyrosine phosphatase A in E. coli

Recombinant plasmid pET30b-PptpA was introduced into competent E. coli BL21(DE3) cells by electroporation (Gene Pulser Electroporation Systems, Bio-Rad). For PtpA production, an overnight pre-culture of transformed cells was added to 250 mL LB medium in an Erlenmeyer flask containing kanamycin. Cells bearing PtpA gene at log phase $\left(\mathrm{OD}_{600}\right.$ of c.a. 0.6 ) were induced by addition of 0.5 mM isoprophyl beta-thiogalactose (IPTG, SigmaAldrich) and kept in a rotary shaker for 4 hours at $37^{\circ} \mathrm{C}, 250 \mathrm{rpm}$. Cells were harvested by centrifugation at 3500 rpm and cell pellet was resuspended in phosphate buffered saline (PBS: $137 \mathrm{mM} \mathrm{NaCl}, 2.7 \mathrm{mM} \mathrm{KCl}, 8 \mathrm{mM} \mathrm{Na} 2 \mathrm{HPO}_{4}$ and $2 \mathrm{mM} \mathrm{KH} 2 \mathrm{PO}_{4}$ ) prior to lysis by 10
cycles of sonication ( 30 seconds sonication and 30 seconds pause). Clear lysate containing PtpA was obtained by centrifugation at $12,000 \mathrm{rpm}, 4^{\circ} \mathrm{C}$ for 30 min .

Protein concentration was determined by bicinchoninic acid (BCA kit, Thermo scientific). Sodium dodecylsufate polyacrylamide gel electrophoresis was performed to show the expression of PtpA in E. coli. Samples were subjected to electrophoresis in $12.5 \%$ polyacrylamide gel.

The activity PtpA was tested for its phosphatase activity against para-nitrophenyl phosphate (p-NPP) as modified from (Mascarello et al., 2013). Reaction mixture contained 6 mM pNPP substrate and $100 \mu \mathrm{M}$ imidazole was added to final volume of $180 \mu \mathrm{~L}$, followed by preincubation at $37^{\circ} \mathrm{C}$ for 5 minutes prior to addition of $20 \mu \mathrm{~L}$ of $\operatorname{PtpA}(1 \mu \mathrm{~g} / \mu \mathrm{L})$. Reaction was allowed to proceed and the resulted para-nitrophenol was measured at 410 nm every 2 minutes for 30 minutes.

### 2.2.2 Inhibition of PtpA by eicosenoic fatty acids

To test the inhibitory effect of eicosenoic fatty acids, PtpA was allowed to hydrolyze substrate para-nitrophenyl phosphate in the absense or presense on various concentrations of trans-2-eicosenoic acid, cis-2-eicosenoic acid, and trans-11-eicosenoic acid, ranging from 0 $\mu \mathrm{M}$ to $30 \mu \mathrm{M}$, for 30 minutes at $37^{\circ} \mathrm{C}$. Lysate of untransformed E. coli BL21 was used as control in each experiments.

The inhibitory effect of eicosenoic fatty acid isomers on the activity of PtpA was depicted as the decrease in absorbance $\left(\mathrm{A}_{410} \mathrm{~nm}\right)$ by the increment concentration of eicosenoic fatty acids. The $\mathrm{IC}_{50}$ values for each eicosenoic fatty acid were calculated with the use of Prism 7 (GraphPad).

### 2.2.3 Docking

Interaction of eicosenoic fatty acids with PtpA that lead to inhibitory of phosphatase activity was analyzed with the Discovery studio (Accelrys, San Diego, CA, USA). The PtpA protein that contains native ligand glycerol (PDB accession number 1U2Q) was used for docking with each of the eicosenoic fatty acids. Ligands, i.e. the eicosenoic fatty acids, were prepared by Chemdraw (Fig. 1B).

## 3. Results

### 3.1 Expression of PtpA in E. coli

$\operatorname{PtpA}_{-2}$ cloned into the $\mathrm{pET30b}$ plasmid, was successfully expressed in E. coli BL21(DE3) under T7 promoter and induced by 0.5 mM isopropyl beta-thiogalactose (IPTG) as described by Studier et al (Studier et al., 1990). PtpA appears as a protein of c.a. 18 kDa (Figure 1A), which is in agreement with another report (Chiaradia et al., 2008). Activity of overexpressed PtpA was assayed by measuring its ability to hydrolyze para-nitrophenyl phosphate ( pNPP ). The released para-nitrophenol gave a typical yellow colour of reaction mixture which is measured spectrophotometrically at 410 nm (Chiaradia et al., 2008).

### 3.2. Inhibition of PtpA with eicosenoic fatty acids

Inhibition of PtpA by trans-2 and cis-2-eicosenoic acids is shown in Figure 2. It is shown that the trans-2 eicosenoic acid ${ }_{2}$ and to a higher extent the cis-2-eicosenoic acid, are capable of inhibiting PtpA.

### 3.3. Docking analysis

Interaction study of PtpA with the cis-2-eicosenoic acid is depicted in Fig. 3. The fatty acid shows interaction with four amino acid residues of PtpA, i.e. Asn ${ }_{14}, \mathrm{Ile}_{15}, \mathrm{Cys}_{16}$ and Arg $_{17}$ with binding energy of $-37.1939 \mathrm{kcal} / \mathrm{mol}$. Strong interaction of PtpA is also found
with trans-2-eicosenoic acid via two potential amino acid residues, i.e. $\mathrm{Thr}_{12}, \mathrm{Gly}_{14}$, and to a lesser degree with $\mathrm{Arg}_{17}$. On the other hand, Ddocking of PtpA with trans-11-eicosenoic acid reveals it to be prone to have weaker interaction in terms of both, the energy and the distance of amino acid residues, which is in agreement with the inhibition assay results. Binding parameters of the tested eicosenoic fatty acids are summarized in Table 1.

Table 1. Energy and binding interaction of co-crystalized Mtb PtpA (1U2Q) with eicosenoic fatty acids

| Compound | cDocker energy <br> $(\mathrm{kcal} / \mathrm{mol})$ | Binding interaction <br> (amino acid residue) |
| :---: | :---: | :---: |
| cis-2-eicosenoic acid | $-37,1939$ | Asn $_{14}, \mathrm{Tle}_{15}, \mathrm{Cys}_{16}, \mathrm{Arg}_{17}$ |
| trans-2-eicosenoic acid | $-33,0076$ | $\mathrm{Thr}_{12}, \mathrm{Gly}_{13}, \mathrm{Arg}_{17}$ |
| trans-11-eicosenoic acid | $-28,3423$ | $\mathrm{Gly}_{13}, \mathrm{His}_{49}$ |

## 4. Discussion

The role of phosphatases in the progress of infection has been of interests of many researchers, since these proteins are identified in various bacterial pathogens, such as Listeria monocytogenes, Mycobacterium tuberculosis, Pseudomonas aeruginosa, Streptococcus agalactiae, Streptococcus pyogenes, Staphylococcus aureus, Salmonella typhimurium, etc. (Sajid et al., 2015). The study areas could include identification of their host interacting partners and be followed by elucidation of the downstream interferences they cause. The next immediate interest, unsuprisingly, is to find a way to inhibit the secreted bacterial phosphatases, and thus prevent their involvement in the development of infections and diseases.

As TB remains a global concern that belongs to one of the main targets in the Sustainbable Development Goal (SGD) in health, prevention of latent TB turns to be one of the focal points, in addition to other TB eradication efforts, such as development of new vaccines, novel drugs, as well as improvement of diagnosis. Our current knowledge on roles of mycobacterial phosphatase effectors that are secreted by Mtb into its host's cells_enroute its degradation pathway has led researchers to explore inhibitors of Mtb phosphatases. The endeavour to seek new potential inhibitors for Mtb virulence proteins and drugs againts TB is nowadays approached by differents strategies. Among those strategies are direct screening of natural compounds, in silico screening of phosphatase inhibitors, and synthesis of novel compounds or modified compounds predicted to be able to inhibit Mtb phosphatases.

The study presented here employs an in silico report that has underlined the potential inhibitory effect of trans-2-eicosenoic fatty acid on Mtb PtpA (Dhanjal et al., 2014). Our data confirmed for the first time that trans-2-eicosenoic fatty acid strongly inhibits PtpA, with an IC $_{50}$ of $11.26 \mu \mathrm{M}$. Interestingly, its cis isomer (cis-2-eicosenoic fatty acid) showed an even stronger inhibition to PtpA, with an $\mathrm{IC}_{50}$ of $8.20 \mu \mathrm{M}$. The ability of these eicosenoic acids to inhibit PtpA is comparable to the other PtpA potential inhibitors, such as chalcone derivatives (Mascarello et al., 2010) and a patented compound (Organization, 2012). In contrast, the trans-11-eicosenoic acid isomer of those compounds showed a much higher $\mathrm{IC}_{50}$ value, i.e. $27.97 \mu \mathrm{M}$ and only slightly inhibited PtpA (Fig. 2). The fact that both trans-2 and cis-2 eicosenoic acids strongly inhibit PtpA, whereas trans-11 does not, shows that the double bond position contributes more to the inhibitory effect of the eicosenoic acids than its cis or trans stereochemistry (Figure 2). It is somehow surprising that a small molecule such as eicosenoic acid, rather than complex inhibitors as many researchers have reported, strongly interacts with PtpA.

## 5. Conclusions

These findings show that both, the cis-2 and trans-2-eicosenoic fatty acids are potential candidates for PtpA inhibitors. A further study is deemed necessary in order to investigate whether these compounds bind specifically to the Mtb's phosphatase by comparing their inhibitory effects on human phosphatases. Additionally, it is also important to employ an assay to prove whether these compounds are capable of preventing Mtb latent infection in vivo.

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## References

Bach H, Papavinasasundaram KG, Wong D, Hmama Z, Av-Gay Y (2008) Mycobacterium tuberculosis Virulence Is Mediated by PtpA Dephosphorylation of Human Vacuolar Protein Sorting 33B. Cell Host Microbe 3: 316-322. https://doi.org/10.1016/j.chom.2008.03.008

Chiaradia LD, Mascarello A, Purificação M, Vernal J, Cordeiro MNS, Zenteno ME, Villarino A, Nunes RJ, Yunes RA, Terenzi H (2008) Synthetic chalcones as efficient inhibitors of Mycobacterium tuberculosis protein tyrosine phosphatase PtpA. Bioorganic Med. Chem. Lett. 18: 6227-6230. https://doi.org/10.1016/j.bmcl.2008.09.105

Dhanjal JK, Grover S, Sharma S, Singh A, Grover A (2014) Structural insights into mode of actions of novel natural Mycobacterium protein tyrosine phosphatase B inhibitors. BMC Genomics 15 Suppl 1: S3. https://doi.org/10.1186/1471-2164-15-S1-S3

Houben RMGJ, Dodd PJ (2016) The Global Burden of Latent Tuberculosis Infection : A Reestimation Using Mathematical Modelling. 5: 1-13. https://doi.org/10.1371/journal.pmed. 1002152

Janssen S, Jayachandran R, Khathi L, Zinsstag J, Grobusch MP, Pieters J (2012) Exploring prospects of novel drugs for tuberculosis. Drug Des. Devel. Ther. 6: 217-224. https://doi.org/10.2147/DDDT.S34006

Korb VC, Chuturgoon AA, Moodley D (2016) Mycobacterium tuberculosis: Manipulator of protective immunity. Int. J. Mol. Sci. 17: https://doi.org/10.3390/ijms17030131

Li W, Xie J (2011) Role of mycobacteria effectors in phagosome maturation blockage and new drug targets discovery. J. Cell. Biochem. 112: 2688-2693. https://doi.org/10.1002/jcb. 23218

Mali PC, Meena LS (2018) Triacylglycerol: nourishing molecule in endurance of Mycobacterium tuberculosis. J. Biosci. 43: 149-154. https://doi.org/10.1007/s12038-018-9729-6

Mascarello A, Chiaradia LD, Vernal J, Villarino A, Guido RVC, Perizzolo P, Poirier V, Wong D, Martins PGA, Nunes RJ, Yunes RA, Andricopulo AD, Av-Gay Y, Terenzi H (2010) Inhibition of Mycobacterium tuberculosis tyrosine phosphatase PtpA by synthetic chalcones: Kinetics, molecular modeling, toxicity and effect on growth. Bioorganic Med. Chem. 18: 3783-3789. https://doi.org/10.1016/j.bmc.2010.04.051

Mascarello A, Mori M, Chiaradia-Delatorre LD, Menegatti ACO, de Monache F, Ferrari F,

Yunes RA, Nunes RJ, Terenzi H, Botta B, Botta M (2013) Discovery of Mycobacterium tuberculosis Protein Tyrosine Phosphatase B (PtpB) Inhibitors from Natural Products. PLoS One 8: 1-12. https://doi.org/10.1371/journal.pone. 0077081
de Oliveira Viana J, Scotti MT, Scotti L (2018) Molecular Docking Studies in Multitarget Antitubercular Drug Discovery. In: Methods in Pharmacology and Toxicology. pp. 187201. https://doi.org/10.1007/7653

Organization WIP (2012) Al 712.

Pieters J, Gatfield J (2002) Hijacking the host: Survival of pathogenic mycobacteria inside macrophages. Trends Microbiol. 10: 142-146. https://doi.org/10.1016/S0966-842X(02)02305-3

Sajid A, Arora G, Singhal A, Kalia VC, Singh Y (2015) Protein Phosphatases of Pathogenic Bacteria: Role in Physiology and Virulence. Annu. Rev. Microbiol. 69: 527-547. https://doi.org/10.1146/annurev-micro-020415-111342

Sens L, de Souza ACA, Pacheco LA, Menegatti ACO, Mori M, Mascarello A, Nunes RJ, Terenzi H (2018) Synthetic thiosemicarbazones as a new class of Mycobacterium tuberculosis protein tyrosine phosphatase A inhibitors. Bioorganic Med. Chem. 26 5742-5750. https://doi.org/10.1016/j.bmc.2018.10.030

Studier FW, Rosenberg AH, Dunn JJ, Dubendorff JW (1990) Use of T7 RNA polymerase to direct expression of cloned genes. In: Methods in Enzymology. pp. 60-89. https://doi.org/10.1016/0076-6879(90)85008-C

Stutz MD, Clark MP, Doerflinger M, Pellegrini M (2018) Mycobacterium tuberculosis: Rewiring host cell signaling to promote infection. J. Leukoc. Biol. 103: 259-268. https://doi.org/10.1002/JLB.4MR0717-277R

Sun-Wada GH, Tabata H, Kawamura N, Aoyama M, Wada Y (2009) Direct recruitment of H+-ATPase from lysosomes for phagosomal acidification. J. Cell Sci. 122: 2504-2513. https://doi.org/10.1242/jcs. 050443

Upadhyay S, Mittal E, Philips JA (2018) Tuberculosis and the art of macrophage manipulation. Pathog. Dis. 76: 1-12. https://doi.org/10.1093/femspd/fty037

Vynnycky E, Fine PEM (2000) Lifetime Risks, Incubation Period, and Serial Interval of Tuberculosis. 152: 247-263.

Wang J, Ge P, Qiang L, Tian F, Zhao D, Chai Q, Zhu M, Zhou R, Meng G, Iwakura Y, Gao GF, Liu CH (2017) The mycobacterial phosphatase PtpA regulates the expression of host genes and promotes cell proliferation. Nat. Commun. 8: https://doi.org/10.1038/s41467-017-00279-z

Wong D, Bach H, Sun J, Hmama Z, Av-Gay Y (2011) Mycobacterium tuberculosis protein tyrosine phosphatase (PtpA) excludes host vacuolar-H+-ATPase to inhibit phagosome acidification. Proc. Natl. Acad. Sci. U. S. A. 108: 19371-6. https://doi.org/10.1073/pnas. 1109201108

World Health Organization (2018) Global Tubercolusis Report.

Zhang D, Lin Y, Chen X, Zhao W, Chen D, Gao M, Wang Q, Wang B, Huang H, Lu Y, Lu Y (2019) Bioorganic Chemistry Docking- and pharmacophore-based virtual screening for the identification of novel Mycobacterium tuberculosis protein tyrosine phosphatase B ( MptpB ) inhibitor with a thiobarbiturate scaffold. 85: 229-239.
https://doi.org/10.1016/j.bioorg.2018.12.038

## Figure Legends

## Figure 1

A

B


Figure 1. A. In each lane, $20 \mu \mathrm{~g}$ of samples were loaded onto $4.5 \%$ focusing gel and $12.5 \%$ resolving polyacrylamide gel for separation. Overexpressed PtpA in Escherichia coli BL21(DE3) appears as a major band at 18 kDa which corresponds to $\operatorname{PtpA}$ (lane 2). Lane 1 is uninduced sample. B. Three isomers of eicosenoic fatty acid structures generated by Chemdraw, tested for their inhibitory effect on Mtb PtpA activity: trans-2-eicosenoic acid, cis-2-eicosenoic acid and trans-11-eicosenoic acid.

## Figure 2



Figure 2. Inhibition study of overexpressed PtpA with increasing concentration of eicosenoic fatty acids (ESA) at $0,8,16$ and $30 \mu \mathrm{M}$. The trans-2-eicosenoic and cis-2-eicosenoic strongly inhibit PtpA, whereas trans-11-eicosenoic acid only slightly inhibits PtpA. All measurements were performed twice and data were normalized to the control E. coli lysate. $\mathrm{IC}_{50}$ values of PtpA inhibition by trans-2, cis-2 and trans-11 eicosenoic acid were calculated with the Prism 7 software (GraphPad) and it was revealed that the $\mathrm{IC}_{50}$ values were $11.26,8.20$ and 27.97 $\mu \mathrm{M}$ for trans-2, cis-2 and trans-11-eicosenoic fatty acid, respectively. ESA: Eicosenoic Fatty Acid.

## Figure 3



Figure 3. A and B: Docking analysis of PtpA inhibition by cis-2-eicosenoic acid. Four amino acid residues of PtpA are in close proximity to the cis-2-eicosenoic acid. C and D: Docking analysis of PtpA inhibition with trans-2-eicosenoic acid. Three amino acids are in close proximity to trans-2-eicosenoic acid.

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