

ChemTech

International Journal of ChemTech Research CODEN (USA): IJCRGG, ISSN: 0974-4290, ISSN(Online):2455-9555 Vol.9, No.12, pp 541-548, 2016

The Response of Dwarf Banana Cavendish Growth and Production under Natural Shade

Muhidin¹*, Gusti R. Sadimantara¹, Sitti Leomo¹, Tresjia C. Rakian¹, Makmur Jaya Arma¹ and Ni Wayan Sri Suliartini¹

Department of Agrotechnology, Faculty of Agriculture, Universitas Halu Oleo Kendari 93232 Southeast Sulawesi Indonesia

Abstract : Banana is important commodity in Indonesia as source of food and energy. Naturally, banana grows in open areas. Planting dwarf banana *cavendish* under natural shade are is limited, because the growth of plant is inhibited due to the low light intensity. Developing dwarf banana *cavendish* that tolerant to shade is important. The tolerant cultivar can be planted on the stand either in the form agroforestry or plantation crops as interplanting. The dwarf banana *cavendish* were collected from various places in Southeast Sulawesi, Indonesia. The research was arranged in a split plot design with three replications. Two factors are considered (i) the existence of natural shade or without shade, and (ii) plant genotype. The parameters observed were the vegetative and the generative character. The results showed that there was different characteristic of the dwarf banana *cavendish* under shade condition base on the threshold of acceptable tolerance. Therefore, it is recommended that dwarf banana accessions selected can be used as source to develop dwarf banana shaded tolerant. **Key word:** Banana, Cavendish, Staple food, Shading.

Introduction

Bananas is member of the family Musaceae^{1,2,3}, generally accepted that is come from Southeast Asia, a biogeographically region including the Malay Peninsula, Indonesia, the Philippines and New Guinea was the primary centre and India was a secondary centre^{4,5,6}. The centre of diversity has been placed in Malaysia or Indonesia⁷. The plants are distributed mainly on margins of tropical rainforests⁸ and now widespread throughout the world.

Bananas are among the most important food crops worldwide^{9,10} and is a major food staple in the tropics¹¹. They include diverse types such as dessert, cooking and roasting bananas^{12,13,14}. The general term 'banana' is used to encompass cultivated varieties of the genus Musa that fall into one of two sub-groups¹⁵, the sweet or dessert banana which makes up approximately 43% of world production, and the cooking banana which makes up approximately 57%.

Banana is source of calorie and mineral, as well as most of the vitamins essential for human nutrition ¹⁶. Bananas are not only rich in carbohydrate and vitamins¹³, including vitamin A ^{17,18,19,20,21,22}, vitamin C ^{18,23}, and B6 ^{24,25,26}, but also rich in mineral such as potassium, magnesium, phosphorus, iron and calcium ^{18,27,28}. They are a good source of dietary fibre¹⁷ and are fat-free. Banana is often the first solid food fed to infant and that as

food supplement ²⁹. It also contains high levels of flavones and antioxidants likes *lutein, zea-xanthin, alpha* and *beta-carotenes* ^{30,31}. Bananas also have several medicinal uses ^{26,32,33}. Banana in the form of banana peel is being use as biosorbent to remove chromium, the toxic heavy metals in waste water ³⁴.

Banana is the best-known tropical fruit and Indonesia is the sixth top banana producer among the top 20 banana producing countries in the world ³⁵. Indonesia produces nearly 5.36 million tons of bananas annually ³⁶. The total per capita consumption in Indonesia is about 22 kg. Indonesia is the ranks 4th in level of banana consumption in the world after the Philippines with 56 kg, Brazil 31 kg and Kenya 24 kg per capita consumption ³⁶. In Indonesia, banana ranks first ³⁵ becoming the most important food and cash crop in Indonesia and grown around the year in the country as a commercial purpose and homestead area for local consumption.

There is growing and enormous potential banana market in Indonesia. The Indonesian population reached 250 million people with banana consumption per capita reached 22 kg. It is predicted that future banana demand in Indonesia will increase sharply due to the increase in population, income and increased diversification in banana processing technology. As with all other crop species, banana production in Indonesia faces major challenges not only from biotic and abiotic stresses, but also from the availability of land to cultivate banana.

The increase banana production in Indonesia faced many obstacles, as a result of the low level implementation in recommended cultivation techniques, the presence of pests and diseases and also limited land that can be used to cultivate. The best alternative to overcome this problem is needed to improve the banana production system that can increase production, productivity and quality of bananas through agroforestry system or intercropping. Intercropping is an essential practice when several economic field crops are competing for the same limited land area ³⁷.

Light has influence on the growth and yield of banana. The light is the main factor affecting on the growth and yield ^{38,39}. There is a close relationship between radiations with net photosynthesis ⁴⁰. Shade directly affects the intensity of the light that falls on the surface of the plant canopy⁴¹. Many physiological process in plants may affected by shade³⁹. It included the rate of photosynthesis, transpiration, respiration and translocation of photosynthate⁴², mineral absorption ⁴³ and root growth^{44,45}.

Shade in plant agroforestry however also has the ability to affect plant growth positively through reduction of loss in soil moisture, retaining of nutrients, suppression of weeds, reduction of soil and plant temperature during the day, raising the night air temperature, reduce water runoff, reduce the decomposition of soil organic matter and enhanced humidity^{16,39,46}. The adaptability of plants to the specified environmental conditions is depending on plant genetic trait ⁴⁷. The ability of plant to cope with shade depends on its ability to continue photosynthesis at light deficit conditions ³⁹. There are two mechanisms adaptation of plant to shading stress are avoidance and tolerance ⁴⁸. The avoidance mechanism is done by getting lighter, while the tolerance mechanism was to get a low light intensity efficiently.

The development of dwarf banana cultivar that tolerant to shade is very important. Because it is naturally to grow and better production, banana requires full sun light or illumination in the open area without shielded. The dwarf banana is the ideal type in agroforestry system ¹⁶, because with the dwarfing banana the population or plant density per unit area will be higher so that the potential production will be higher also. Therefore it is necessary screening to obtain the dwarf banana cultivar that shade tolerant and have a high potential production. The present study has provided an assessment of the banana's characteristics towards the existence of shade or low light intensity.

Experimental

The research was arranged in a split plot design with three replications. Two factors are considered (i) the existence of shade or without shade in as main plots, and (ii) the dwarf banana genotype or cultivar in subplots. Observations were made on the vegetative and generative parameter:

- 1. The vegetative variables include plant height and number of leaves that were measured every month.
- 2. The leaf characters include leaf length and leaf width on average, measured from one plant.
- 3. The generative characters include time of flowering, flowering age, harvesting age and weight of the harvest.

The observed data were then analyzed using analysis of variance. Duncan's Multiple Range Test (DMRT) at 95% confidence level was performed to see the effect of different soil conditions on vegetative and generative components.

Results and Discussion

A. Effect of Natural Shade of Vegetative Character

The results showed that there were significant differences in the parameter of plant height, leaf length and leaves width (Table 1 and 2). Based on the data it indicates that shade treatment can increase the characters of plant height, leaf length and leaves width. On the parameter number of leaves, there are no significant differences between plant growing under natural shade and that of without natural shade. The number of leaves usually almost identical between plants growing under natural shade, as well as with plants growing without natural shade (Table 2).

Table 1. The eeffect of natural shade against plant high and leaf length on the plant age up to 10 months after the transfer (MAT)

Age	Plant height (cm)		Leaf length (cm)	
(month)	No natural shade	Natural shade	No natural shade	Natural shade
1	14.67 ^a	24.67 ^b	21.39 ^a	30.29 ^b
2	30.28 ^a	39.28 ^b	30.92 ^a	40.32 ^b
3	40.72 ^a	53.72 ^b	48.37 ^a	60.47 ^b
4	60.70 ^a	80.70 ^b	53.20 ^a	71.40 ^b
5	71.20 ^a	83.20 ^b	69.80 ^a	81.70 ^b
6	82.10 ^a	97.10 ^b	79.30 ^a	92.60 ^b
7	96.40 ^a	106.40 ^b	107.80 ^a	118.00 ^b
8	109.30 ^a	119.30 ^b	142.00 ^a	151.70 ^b
9	117.50 ^a	129.50 ^b	162.00 ^a	173.00 ^b
10	118.50 ^a	130.50 ^b	164.00 ^a	179.00 ^b

Note: Means in the same line suffixed with different letters are different at 5% levels of significance according to DMRT.

Table 2. The effect of natural shade against plant number of leaves	and Leaves Width on the plant age
up to 10 months after the transfer (MAT)	

Age	Number of leaves		Leaves Width (cm)	
(month)	No natural shade	Natural shade	No natural shade	Natural shade
1	3.19 ^a	3.39 ^b	12.20 ^a	16.40 ^b
2	4.40 ^a	4.10 ^a	15.54 ^a	21.64 ^b
3	6.00 ^a	5.90 ^a	18.20 ^a	22.00 ^b
4	6.92 ^a	7.12 ^a	25.70 ^a	32.90 ^b
5	7.80 ^a	7.40 ^b	34.80 ^a	39.92 ^b
6	8.20 ^a	8.70 ^b	48.50 ^a	56.31 ^b
7	9.20 ^a	8.80 ^b	57.60 ^a	66.36 ^b
8	9.60 ^a	9.80 ^a	69.40 ^a	76.50 ^b
9	10.20 ^a	10.40 ^a	73.00 ^a	81.90 ^b
10	11.20 ^a	11.48 ^a	76.54 ^a	82.93 ^b

Note: Means in the same line suffixed with different letters are different at 5% levels of significance according to DMRT.

In general, it appears that the natural shade can increase the plant height, leaf length and leaves width. While the parameter number of leaves there is no definite pattern. It is supposedly that in a higher level of shade, the plant has undergone etiolating so the parameter of plant height, leaf length and leaves width becomes higher.

B. Effect of Natural Shade of Generative Character

Based on the generative character appears that the harvesting time in natural shade conditions, generally longer than that the harvesting time on condition without shade (Table 3). In general, it appears that in the shaded conditions, the production parameters has decreased, especially in the character of total production, the number of combs, number of fruit in each comb and the banana grain size. Nevertheless, there is a tendency increasing in the number of bunches and bunches length increased, although the size and weight of fruit per banana tends to decrease.

No.	Character	No Natural Shade	Natural shade	± (%)
1	Time of flowering (month)	9.2 ^a	9.8 ^b	6.52
2	Time of ripening (month)	11.00 ^a	11.6 ^b	5.45
3	Bunch weight (kg/bunch)	14.26 ^a	13.80 ^b	3.23
4	Total comb (Fruit)	7.8 ^a	8.1 ^b	3.85
5	Bunches length (cm)	59.8 ^a	68.72 ^b	14.92
6	Banana amount for each comb (fruit)	17.2 ^ª	16.3	5.23
7	Total combs of bunch	7.8 ^a	8.2 ^b	5.13
8	Fruit length (cm)	13.5 ^a	13.1 ^b	1.48
9	Fruit width (cm)	3.2 ^a	3.0 ^b	3.13
10	Fruit diameter (cm)	31.3 ^a	28.3	9.58
11	Banana amount for each bunch (fruit)	138.3 ^a	134.5 ^a	2.75
12	Banana weight (g fruit ⁻¹)	103.2 ^a	92.8	10.08
13	Weight flesh (g)	62.6 ^a	54.7	12.62
14	Edible portion of fruit (%)	60.45 ^a	59.00 ^a	2.40

Table 3.	The effect of natural shade on severa	l generative characters	of dwarf banana Cavendish
----------	---------------------------------------	-------------------------	---------------------------

Note: Means in the same line suffixed with different letters are different at 5% levels of significance according to DMRT.

In general, based on Table 3 it appears that there is a decline in the production of dwarf banana Cavendish under the natural shade condition. Decreasing or increasing of the production characters that occur on average only 6 percent and still below the threshold of decline that can be tolerated by 10 percent. Therefore the dwarf banana Cavendish accessions tested can be used as a source of germplasm for banana cultivation in agroforestry system or in polyculture as intercropping patterns.

Light is a key factor in the ultimate growth and production because it supplies the energy needed to fix carbon ⁴⁹. Most of the plant responses to shading are with lower dry matter production, photosynthetic retention in the stem, longer spear development and bigger or thinner leaves ^{50,51}The change in microclimate due to shade levels caused clear differences between growth and plant development ^{50,52,53} and plant height was higher under shaded conditions compared that of without shading.

Plants grown under low light levels have shown more apical dominance than those grown in high light environment resulting in taller plants under shade ⁵⁴. Shading produced taller plants due to shoot growth as they looked for light, indicating the activation of some phototropism responses ⁵⁵, to modify plant leaf distribution in order to limit mutual shading. Enhancing plant height under shade represents a cost to resource acquisition through reduction in either root or leaf allocation ^{50,56,57,58,59}. Plants responses to light include a variety of adaptations at physiological levels ⁶⁰. Such responses are translated into alteration of growth rate and plant architecture ⁶¹. The architecture of plant canopy influences the interception, absorption and scattering of solar radiation ⁶². Reduced light quantity and/or quality also can cause accelerated stem elongation and branch reduction ⁶³.

Irradiation is an important factor affecting crop photosynthesis, development and yield⁶⁴. It is also an important factor helping plants to reach enough light⁵⁰. Similar results also found in faba bean when exposure to shade⁵⁶. Shading is reduced stomatal conductance and photosynthesis rate in *Salvia sclarea*⁵⁰, myrtle⁶⁵ and banana⁶⁶, therefore a reduction in the rate of growth^{67,68}.

Many physiological processes in plants are affected by irradiance, which is one of the most important

environmental factors affecting plant survival, growth, reproduction and distribution ^{50,69}. The amount of light reaching inside the canopy and absorbed by the plant changes with plant density. Yield reduction by shading will depends upon crop species as well as the degree of shading. Shading occurs mainly due to dense plant population, intercropping, planting geometry and excessive vegetative growth, which affects the crop performance through reducing photosynthetic capacity of plant ⁷⁰.

Limited land need to look for a solution through the use of land between tree crop forestry or perennial crops such as rubber, coconut and others to grow dwarf banana in agroforestry or polyculture farming system. Agroforestry has been designed to increases the yield of the land, combines the production of crops and forest plants simultaneously or sequentially on the same unit of land^{71,72}, to obtain economic, environmental, ecological, and cultural benefits⁷³.

Agroforestry options may provide a means for diversifying production systems and increasing the sustainability of smallholder farming systems, through growing food crops in the forests and establishing tree crop production systems on arable lands⁷⁴.

Conclusion

In conclusion it appears that there is significant different on the vegetative character between the dwarf bananas Cavendish that growth under the shade condition compare that of without shade. In the generative character tested, there is decrease or increase in several generative characters, but still below the threshold of acceptable tolerance decline by 10 percent. Therefore accessions selected plants can be used to source the development of banana plants in shaded conditions.

Acknowledgements

The research was supported by the Directorate General of Higher Education, Ministry of Research, Technology and Higher Education of the Republic of Indonesia.

References

- 1. Kress, W.J. and Specht C.D. 2006. The evolutionary and biogeographic origin and diversification of the tropical monocot order Zingiberales. Aliso 22: 621-632.
- 2. Liu, A.Z., Kress, W.J and Li D.Z. Li. 2010. Phylogenetic Analyses of the Banana Family (Musaceae) Based on Nuclear Ribosomal (ITS) and Chloroplast Evidence. Taxon 59 (1) : 20-28.
- 3. Heslop-Harrison, J. S and Schwarzacher T. 2007. Domestication, Genomics and the Future for Banana. Annals of Botany. 100 : 1073–1084
- 4. Simmonds, N.W. and Shepherd K. 1955. The taxonomy and origins of the cultivated banana. J. Linn. Soc. (Botany) 55:302-312.
- 5. Stover, R.H., and Simmonds, N.W. 1987. Bananas. Third edition. John Wiley and Sons, New York.
- 6. Robinson, J.C and Sauco V.G. 2010. Bananas and Plantain. 2nd Edition. CAB International Publisher. United Kingdom's. 320p.
- 7. Daniells JW, Jenny C, Karamura DA, Tomekpe K, Arnaud E, Sharrock S. (eds). 2001. Musalogue: A catalogue of Musa germplasm. Diversity in the genus Musa. Montpellier, France: INIBAP.
- 8. Wong C, Kiew R, Argent G, Set O, Lee SK, Gan YY. 2002. Assessment of the validity of the sections in Musa (*Musaceae*) using AFLP. Annals of Botany 90: 231–238.
- 9. Nelson, S.C. Ploetz, R.C., Kepler, A.K. 2006. Musa Species (banana and plantain). In: CR Elevitch, ed. Species Profiles for Pacific Island Agroforestry. Permanent Agricultural Resources, Holualoa, Hawai'i
- 10. Aurore G, Parfait B, Fahrasmane L. 2009. Bananas, raw materials for making processed food products. Trends Food Sci Technol. 20: 78-91.
- 11. Hölscher, D. Dhakshinamoorthy, S. Alexandrov, T. Becker, M. Bretschneider, T. Buerkert, A. Crecelius, AC. Waele, DD. Elsen, A. Heckel, DG. Heklau, H. Hertweck, C. Kai, M. Knop, K. Krafft, C. Maddula, RK. Matthäus, C. Popp, J. Schneider, B. Schubert, US. Sikora, RA. Svatos, A and Swennen RL. 2014. Phenalenone-type phytoalexins mediate resistance of banana plants (Musa spp.) to the burrowing nematode Radopholus similis. *PNAS* vol. 111 (1) p. 105-110
- 12. Swennen, R., and Vuylsteke, D. 1987. Morphological taxonomy of plantain (Musa cultivars AAB) in

West Africa. Banana and Plantain Breeding Strategies. Proceedings of an International Workshop Held at Cairns, Australia, 13-17 October 1986 (G.J. Persley and E.A. De Langhe, eds.). Australian Centre for International Agricultural Research, ACIAR Proceedings 21: 165-171.

- Sagi, L., May, GD. Remy, S and Swennen, R. 1998. Recent Development in Biotechnological Research on Banana (Musa spp).Biotechnology and Genetic Engineering Reviews. Vol 15 April 1998. P313-327
- Valmayor, R.V., Jamaluddin, S.H., Silayoi, B., Kusumo, S., Danh, L.D., Pascua, O.C., Espino, R.R.C. 2000Banana Cultivar Names and Synonyms in Southeast Asia. International Network for the Improvement of Banana and Plantain - Asia and the Pacific Office, Laguna, Philippines.
- 15. Pillay, M.A., Tenkouano, A., Ude, G., Ortiz, R. 2004. Molecular characterization of genomes in Musa and its applications. Chapter 23. In: Jain, SM. Swennen, R (eds.) Banana Improvement: Cellular, Molecular Biology and Induced Mutations. Science Publishers Inc., Enfield (NH), USA.
- 16. Muhidin, Leomo, S and. Rakian TC. 2015. Pisang Kate : Sumber Pangan dan Energi yang Terabaikan. Unhalu Press Kendari, Indonesia (In Indonesian)
- 17. Blades, B. L., Dufficy, L., Englberger, L., Daniells, J. W., Coyne, T., Hamill, S., Wills, R. B. H. 2003. Bananas and plantains as a source of provitamin A. Asia Pacific Journal of Clinical Nutrition (12) : 9-9
- 18. Wall, M.M. 2006. Ascorbic acid, vitamin A, and mineral composition of banana (Musa sp.) and papaya (*Carica papaya*) cultivars grown in Hawaii. Journal of Food Composition and Analysis 19 : 434–445
- 19. Davey, MW, Van den Bergh, I, Markham, R, Swennen, R and Keulemans, J. 2009. Genetic variability in Musa fruit provitamin A carotenoids, lutein and mineral micronutrient contents. Food Chemistry 115(3):806-813
- 20. Pereira, L.C. 2009. Provitamin A Carotenoids Content in Musa Fruit: Genetic and Ecological Distribution and Rapid Pre-Screening. MSc Thesis, Institut Supérieur d'Agriculture de Lille, France.
- Englberger, L. 2012. Revisiting the vitamin A fiasco: going local in Micronesia. p.126-133. In: Burlingame, B. and S. Dernini (eds.). Proceedings of Biodiversity and sustainable diets united against hunger, 3–5 November 2010. Sustainable diets and biodiversity: Directions and solutions for policy, research and action. FAO, Rome, Italy.
- 22. Ekesa. B.N., Miroir, C. Blomme, G. Van den Bergh I and Davey, M.W. 2013. Retention of provitamin A carotenoids during post-harvest ripening and processing of three popular Musa cultivars in South-Western Uganda. Acta Horticulturae 986: 319-330.
- Kumar, G.V., Kumar, K. A., Patel, G.R.R., and Manjappa, S. 2013. Determination of vitamin C in some fruits and vegetables in Davanagere city, (Karanataka) India. Int. J. of Pharm. & Life Sci. (IJPLS) 4 (3): 2489-2491
- 24. Adeyemi, O. S. and Oladiji, A. 2009. Compositional changes in banana (*Musa* ssp.) fruits during ripening. African Journal of Biotechnology 8 (5): 858-859.
- 25. Abiodun-Solanke, A.O and Falade, K.O. 2010. A Review of the uses and methods of processing banana dan plantain (*Musa* spp) into storable food products. J. Agris Res and Dev 9 (2) : 85-166
- 26. Kumar, K.P.S., Bhowmik, D. Duraivel, S and Umadevi, M. 2012. Traditional and Medicinal Uses of Banana. Journal of Pharmacognosy and Phytochemistry 1 (3) : 51-63
- 27. Anhwange, B.A., Ugye T.J and Nyiaataghe, T.D. 2009. Chemical Composition of *Musa sepientum* (Banana) Peels. EJEAFChe, 8 (6) : 437-442
- 28. Okorie, D.O., Eleazu, C.O. and Nwosu, P. 2015. Nutrient and Heavy Metal Composition of Plantain (*Musa paradisiaca*) and Banana (*Musa paradisiaca*) Peels. J Nutr Food Sci 5(3) : 1-3
- 29. Ekesa, B.N. 2014. Vitamin A Banana/Plantain- Biofortification. The 2nd Global Conference on Biofortification: Getting Nutritious Foods to People. March 31 to 1 April 2014. Kigali. Rwanda.
- 30. Englberger. L., Schierle, J. Aalbersberg, W. Hofmann, P. Humphries, J and Huang, A. 2006. Carotenoid and vitamin content of Karat and other Micronesian banana cultivars. International Journal of Food Sciences and Nutrition 57(5-6): 399-418.
- Ekesa, B.N., Poulaert, M. Davey, MW. Kimiywe, J. Van den Bergh, I. Blomme, G. and Dhuique-Mayer, C. 2012. Bioaccessibility of provitamin A carotenoids in bananas (Musa spp.) and derived dishes in African countries. Food Chemistry 133: 1471–1477.
- 32. Pushpangadan, P., Kaur, J and Sharma, J. 1989. Plantain or Edible Banana (*Musa X paradisica* Var *Sapientum*) Some Known Folk Uses In India. Ancient Science of Life 9 (1) :20-24
- Venkatesh., Krishna, V. Girish, K.K. Pradeepa, K. And Kumar, S.S.R. 2013. Antibacterial Activity of Ethanol Extract of *Musa paradisiaca* Cv. Puttabale And *Musa Acuminate* Cv. Grand Naine. Asian J. Pharm Clin Res, Vol 6, Suppl 2 : 169-172

- 34. Nirmal, X.M and Ragupathi, T. 2015. Experimental Study on Reduction of *Chromium* Conten from Indsutrial Effluent Using Banana Peel. Int. J. ChemTech Res. 8 (6) : 725-731.
- 35. Yusnita., E. Daniel and Hapsoro, D. 2015. In Vitro Shoot Regeneration of Indonesian Banana (*Musa* spp) cv Ambon Kuning and Raja Bulu, Planlet Acclimatization and Field Performance. Agrivita 37 : 51-58
- 36. FAOSTAT. 2014. Food and Agricultural commodities production / Countries by commodity. Food and Agriculture Organization Of The United Nations Statistics Division. Rome. Italy
- Abd Al-Lateef, E.M., Abd El-Salam, M.S. El-Habbasha, S.F. and Ahmed, M.A. 2015. Effect of Maize – Cowpea Intercropping on Light Interception, Yield and Land Use Efficiency. Int. J. ChemTech Res. 2015, 8 (6), pp 556-564.
- 38. Fischer RA., 1975. Yield Potential in Dwarf Spring Wheat and the Effect of Shading. Crop Science. 15:607-613
- Muhidin. Jusoff, K. Elkawakib, S. Musa, Y. Kaimuddin. Meisanti. Sadimantara, G.R. and Baka, L.R. 2013. The Development of Upland Red Rice under Shade Trees. World Applied Sciences Journal 24 (1): 23-30, 2013
- 40. Webster, C.C and Wilson PN. 1980. Agriculture in The Tropics 2nd. Logman, London. 371p.
- 41. Man JE, Curry GL, Michele D.W and Baker DN. 1980. Light Penetration in a Row-Crop with Random Plant Spacing. Agronomy Journal. 72:131-139.
- 42. Struik PC and Deinum B. 1982. Effect of Light Intensity after Flowering on the Productivity and Quality of Silage Maize. Netherlands Journal of Agriculture Science, 30: 297-316.
- 43. Marschner H. 1995. Mineral Nutrition of Higher Plants. Academics Press Inc. San Diego. 889p.
- 44. Wong, C.C and Wilson J.R. 1980. Effect of Shading on the Growth and Nitrogen Content of Grain Panicle and Siratro in Pure and Mixed Swards Defoliated at Two Frequencies. *Aus. J. Agric. Res.* 31:269-289
- 45. Stoskopf. N.C. 1981. Understanding Crop Production. Reston Publishing Company Inc. Reston Virginia. 433 p.
- 46. Strigter CJ. 1984. Shading: A Traditional Method of Microclimate Manipulation. Netherlands Journal of Agriculture, 32: 81-86.
- 47. Mohr, H and Schoopfer, P. 1995. Plant physiology. Translator Gudrum and David W. Lawlor, Springer-Verlag.NY. 629p.
- 48. Levitt J. 1980. Responses of plants to environmental stresses, Vol. 2. Water, radiation, salt and other stresses. New York: Academic Press, p.93–128
- 49. Maffei, M. 2003. Dietary Supplements of Plant Origin: A Nutrition and Health Approach. Taylor and Francis Group. CRC Press. London. 256p
- 50. Kumar, R., Sharma, S and Pathania, V. 2013. Effect of shading and plant density on growth, yield and oil composition of clary sage (*Salvia sclarea* L.) in north western Himalaya, Journal of Essential Oil Research, 25:1, 23-32.
- 51. Dapont, E.C. Da Silva, JB and Alves, CZ. 2016. Initial Development Of Açaí Plants Under Shade Gradation. Rev. Bras. Frutic., 38 (2) : 1-22
- 52. Holcman, E., Sentelhas, PC. 2013. Bromeliads production in greenhouses associated to different shading screens. Horticultura Brasileira 31: 386-391
- 53. Fadil, M., Farah, A., Ihssane, B., Haloui, T., Lebrazi, S., Zghari, B., and Rachiq, S. 2016. Chemometric investigation of light-shade effects on essential oil yield and morphology of Moroccan *Myrtus communis* L. SpringerPlus 5:1062
- 54. Moniruzzaman, M., Islam, M.S. Hossain, M.M. Hossain, T and Miah, M.G. 2009. Effects of shade and nitrogen levels on quality Bangladhonia production. Bangladesh J. Agric.Res., 34 : 205–213.
- 55. Takemiya, A. Inouea, S. Doi, M. Kinoshita, T and Shimazaki, K. 2005. Phototropins promote plant growth in response to blue light in low light environments. Plant Cell, 17 : 1120–1127.
- 56. Nasrullahzadeh, S., Ghassemi-Golezani, K. Javanshir, K. Valizade, K. and Shakiba, MR. 2007. Effects of shade stress on ground cover and grain yield of faba bean (*Vicia faba* L.). J. Food Agric. Envt., 5, 337–340.
- 57. Kumar, R., Sharma, S. Ramesh, K. and Singh, B. 2013. Effects of shade regimes and planting geometry on growth, yield and quality of the natural sweetener plant stevia (*Stevia rebaudiana* Bertoni) in north-western Himalaya. Archives of Agronomy and Soil Science Vol. 59, Issue 7: 963-979.
- 58. Modrzyński, J., Chmura, DJ. and Tjoelker, MG. 2015. Seedling growth and biomass allocation in relation to leaf habit and shade tolerance among 10 temperate tree species. Tree Physiology. Volume 35, Issue 8 : 879-893

- 59. Fazlioglu, F., Al-Namazi, A and Bonser, SP. 2016. Reproductive efficiency and shade avoidance plasticity under simulated competition. Ecology and Evolution 6(14): 4947–4957.
- 60. Wigington, J. R. and McMillan, C.1979. Chlorophyll composition under controlled light conditions as related to the distribution of seagrass in Texas and the U. S. Virgin Islands. Aquat. Bot. 6:171-184.
- 61. Backman, T. W. and Barilotti, D. C. 1976. Irradiance reduction: Effects on shading crops of the eelgrass (*Zostera marina*) in a coastal lagoon. Mar. Biol. 43:33-40.
- 62. Christopher, J. K., John, M. N. and Smith, T. G. 1998. Measurement of leaf orientation, light distribution and sunlit leaf area in a boreal aspen forest. Agriculture and Forest Meteorol. 91:127-148.
- 63. Deregibus, V. A., Sanchez, R. A., Casal, J. J. and Trlica, M. J. 1985. Tillering responses to enrichment of red light beneath the canopy in a humid natural grassland. J. Appl. Ecol. 22:199-206.
- 64. Zhao, D. and Oosterhuis, D. 1998. Cotton responses to shade at different growth stages: Nonstructural carbohydrate composition. Crop Sci. 38:1196-1203.
- 65. Mendes, M. M., Gazarine, L.C. and Rodrigues, M.L. 2001. Acclimation of *Myrtus communis* to contrasting Mediterranean light environments effects on structure and chemical composition of foliage and plant water relations. Environ. Exp. Bot., 45, 165–178.
- 66. Israeli, Y., Plaut, Z and Schwartz, A. 1995. Effect of shade on banana morphology, growth and production. Sci. Hort.62 : 45–56.
- 67. Corre, W.J. 1983. Growth and morphogenesis of sun and shade plants. II. The influence of light quality. Acta Bot. Neerl., 32 : 185–202.
- Dold, C., Charles, S. Luis, P. Joachim, H. 2008. Musa in Shaded Perennial Crops Response to Light Interception. Conference on International Research on Food Security, Natural Resource Management and Rural Development. University of Hohenheim, Tropentag. October 7-9, 2008
- 69. Keller, P and Luettge, U. 2005. Photosynthetic light-use by three bromeliads originating from shaded sites (Ananas ananassoides, Ananas comosus cv. Panare) and exposed sites (Pitcairnia pruinosa) in the medium Orinoco basin, Venezuela. Biol. Plant., 49: 73–79.
- 70. Noggle, G. R and Fritz, G.J. 1979. Introductory Plant Physiology, p. 688. Prentice Hall of India Pvt. Ltd., New Delhi.
- 71. King. K.F.S. 1979. Agroforestry and The Utilisation of Fragile Ecosystems. Journal of forest Ecology and Management, 2 : 161–168.
- 72. Smith. N.J.H., Falesi, I.C. Alvim, F.T. and Serrao, E.A.S. 1996. Agroforestry Trajectories Among Smallholder in The Brazilian Amazon : Innovation and Resiliency in Pioneer and Older Settled Area. Journal of Ecological Economic, 18 : 5-27
- 73. Chauhan. S. K., Dhillon, WS. Singh, N and Sharma, R. 2013. Physiological Behaviour and Yield Evaluation of Agronomic Crops under Agri-horti-silviculture System. International Journal of Plant Research 3(1): 1-8
- 74. Kumar. B.M., 2006. Agroforestry: The New Old Paradigm for Asian Food Security (Review). Journal of Tropical Agriculture 44 (1-2): 1-14.
